

Optogenetics in Neurology and Neuroscience

Taruna Ikrar

Hasanuddin University, Makassar, Indonesia.

Brain Circulation Institute of Indonesia (BCII), Indonesia.

Department of Anatomy and Neurobiology, University of California, Irvine, USA.

Keywords— optogenetics, neurology, neuroscience.

Copyright©2017. Published by UNSYSdigital. All rights reserved.
DOI: [10.21535/pcs.v3i1.938](https://doi.org/10.21535/pcs.v3i1.938)

OPTOGENETICS permit researchers and scientists to make a light sparkling on cellular elements and neurons, especially on distal site of axon terminals, also hyperpolarize or depolarize them whom express opsin. Techniques of optogenetics in neurovascular systems need three main strategies. First, we develop an opsin to get the desirable neuronal effects. Second, we express the opsin in the desirable cellular elements. Third, we deliver light into the opsin [1],[2].

One of applications of optogenetics techniques is researches about multiregional-neuronal circuits in the brain. They are hippocampus, cerebral cortex, nucleus accumbens, and striatum. Many studies in non-human primate and rodent have proven that optogenetic neuromodulation can potentially be utilized to manage multidisorders in neurological and neuroscience fields [3], such as: motoric dysfunction in Parkinsonian animals [4] and ameliorating visual disabilities in blind rodents that genetically modified [5].

Scientists have utilized optogenetics human retina preparations in vitro to enhance light sensitivity to the prior light-insensitive photoreceptors of blind patients. Optogenetics also have important roles in neuromodulation therapies [6],[7].

A paragon pharmacological perspective in epilepsy would target just brain regions in charge of seizures events, which impossible with systemically delivered antiepileptic medications. Moreover this treatment ought not meddle with normal physiological capacity. Since seizures are discontinuous, an imperative development would be accomplished by building up effective strategies for the fast and reversible concealment of action in a confined region of the brain. An approach to smother seizure activity 'on interest' is to photo-activate light-sensitive ion channels and transporters that have been revealed in neurons. There are two general optogenetic strategies scientist could use to hinder seizures. First, we express halorhodopsin (an inhibitory opsin) in excitatory neurons to smother their excitability and diminish

output. Second, we express a suitable opsin in groups of interneurons to manage their firing in a way that would bring about increased inhibition of main neurons [8]-[10].

Researches in epilepsy-related optogenetic have concentrated on expressing halorhodopsin in paramount neurons to smother seizure commotion. Moreover, opsin hindrance or actuation of interneurons may likewise be viable and have begun to attract attention the previous couple of years. A few clusters have utilized channel rhodopsin-2 communicated in interneurons to control seizure commotion in different in vitro and in vivo epilepsy model [11]-[13].

In the long period, an interneuron based optogenetic perspective will be very useful to repair the inhibition-excitation balance with a minimal disruption of the neuronal network [14].

REFERENCES

- [1] Zhang F, Gradinaru V, Adamantidis AR, et al. Optogenetic interrogation of neural circuits: technology for probing mammalian brain structures. *Nat Protoc.* 2010;5(3):439–456. [CrossRef](#)
- [2] Rossi MA, Calakos N, Yin HH. Spotlight on Movement Disorders: What optogenetics has to offer. *Mov Disord.* 2015;30(5):624–631. [CrossRef](#)
- [3] Tonnesen J. Optogenetic cell control in experimental models of neurological disorders. *Behav Brain Res* 2013; 255:35–43. [CrossRef](#)
- [4] Kravitz AV, Freeze BS, Parker PR, Kay K, Thwin MT, Deisseroth K, et al. Regulation of parkinsonian motor behaviours by optogenetic control of basal ganglia circuitry. *Nature* 2010;466:622–6. [CrossRef](#)
- [5] Doroudchi MM, Greenberg KP, Liu J, Silka KA, Boyden ES, Lockridge JA, et al. Viroly delivered channelrhodopsin-2 safely and effectively restores visual function in multiple mouse models of blindness. *Mol Ther: J Am Soc Gene Ther* 2011;19:1220–9. [CrossRef](#)
- [6] Busskamp V, Duebel J, Balya D, et al. Genetic reactivation of cone photoreceptors restores visual responses in retinitis pigmentosa. *Science.* 2010;329(5990):413–417. [CrossRef](#)
- [7] Williams JC, Denison T. From optogenetic technologies to neuromodulation therapies. *Sci Transl Med.* 2013;5(177):176-177. [CrossRef](#)
- [8] Shiri Z, Manseau F, Levesque M, Williams S, Avoli M. Interneuron activity leads to initiation of low-voltage fast-onset seizures. *Ann Neurol* 2015;77:541–6. [CrossRef](#)
- [9] Yekhlief L, Breschi GL, Lagostena L, Russo G, Taverna S. Selective activation of parvalbumin- or somatostatin-expressing interneurons triggers epileptic seizure-like activity in mouse medial entorhinal cortex. *J Neurophysiol* 2015;113:1616–30. [CrossRef](#)
- [10] Wykes RC, Kullmann DM, Pavlov I, Magloire V. Optogenetic approaches to treat epilepsy. *Journal of Neuroscience Methods* 2016;260:215–220. [CrossRef](#)

Corresponding author: Taruna Ikrar (email: dr.ikrar.phd@gmail.com).

This paper was submitted on March 15, 2017; revised on April 15, 2017; and accepted on April 15, 2017.

- [11] Krook-Magnuson E, Armstrong C, Oijala M, Soltesz I. On-demand optogenetic control of spontaneous seizures in temporal lobe epilepsy. *Nat Commun* 2013;4:1376. [CrossRef](#)
- [12] Ellender TJ, Raimondo JV, Irkle A, Lamsa KP, Akerman CJ. Excitatory effects of parvalbumin-expressing interneurons maintain hippocampal epileptiform activity via synchronous afterdischarges. *J Neurosci: Off J Soc Neurosci* 2014;34:15208–22. [CrossRef](#)
- [13] Ledri M, Madsen MG, Nikitidou L, Kirik D, Kokaia M. Global optogenetic activation of inhibitory interneurons during epileptiform activity. *J Neurosci: Off J Soc Neurosci* 2014;34:3364–77. [CrossRef](#)
- [14] Wykes RC, Kullmann DM, Pavlov I, Magloire V. Optogenetic approaches to treat epilepsy. *J Neurosci Methods* 2016;260:215–220. [CrossRef](#)